protein genespin

NEW PRODUCT _Green Line

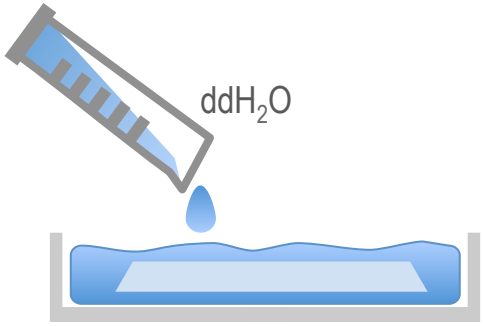
In classical protein staining protocols using Coomassie Brilliant Blue G-250, solutions with high contents of toxic and flammable organic solvents (Methanol, Ethanol or 2-Propanol) and acetic acid are used for fixation, staining and destaining of proteins in a gel after SDS-PAGE. To speed up the procedure, heating the staining solution in the microwave oven for a short time is frequently used. This usually results in evaporation of toxic or hazardous Methanol, Ethanol or 2-Propanol and a strong smell of acetic acid in the lab which should be avoided due to safety considerations.

GeneSpin Coomassie Stain is **a ready-to-use non hazardous solution** for staining proteins on polyacrylamide gels in 20-25 minutes. GeneSpin Coomassie Stain **does not require addition of methanol and acetic acid for destaining** and produces blue bands on a clear background.

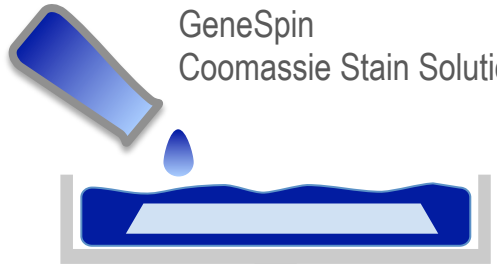
> Coomassie Stain

cat.no	*amount*	*note*
STS-CS 500	2 x 250ml (500mL)	methanol, ethanol, acetic acid free
STS-CS 1000	4 x 250ml (1L)	methanol, ethanol, acetic acid free

ADVANTAGES:
- Staining in aqueous solution (no special handling or fume hood requirements) in 20-25 minutes.
- Visibility of bands while gel is in the stain.
- No solvent waste problems.



1. About 100ml of bidistilled water is added to the gel and heated in the microwave oven for 45 seconds. (avoid boiling). The box with the gel is then placed on a shaker for 3-5 min. Repeat twice this step with fresh water.



2. Cover the gel in the box with GeneSpin CS solution and heat in the microwave for 30 sec. without boiling. Placed the box on a shaker for finishing the staining.

3. **After 1 minute**, protein bands can be observed, **after 15-30 minutes** the staining is strong enough in most cases.

4. Pour off CS Solution and add 50-100 ml bidistilled water to further destain the light blue background of the gel on ashaker

5. The gel can be scanned, photographed or dried for long-term storage

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